

1 Human Viruses in Water 39
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Chapter 3

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11
Enteric Hepatitis Viruses

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23
Background

25

25 The term “jaundice” was used as early as in the ancient Greece when Hippocrates
27 described an illness probably corresponding to a viral hepatitis. However, it was
not until the beginning of the twentieth century when a form of hepatitis was
29 associated to an infectious disease occurring in epidemics and the term “infectious
31 hepatitis” was established. Later on in the early 1940s two separate entities were
defined: “infectious” and “serum” hepatitis, and from 1965 to nowadays the
33 different etiological agents of viral hepatitis have been identified. Although all viral
35 hepatitis are infectious the aforementioned terms refer to the mode of transmission,
corresponding the “infectious” entity to those hepatitis transmitted through the
37 fecal-oral route and the “serum” hepatitis to those transmitted parenterally. Thus,
the infectious or enteric hepatitis include two types: hepatitis A and hepatitis E,
which will be reviewed here.

39
Hepatitis A41
*Natural course and epidemiology of hepatitis A*43
Hepatitis A infection may develop asymptotically. This type of subclinical in-
fection is most common among young children (under 5), while in older children

1 and in the adulthood the infection usually proceeds with symptoms (Previsani et
2 al., 2004). In this latter case, the clinical course of hepatitis A is indistinguishable
3 from that of other types of acute viral hepatitis. The clinical case definition for
4 hepatitis A is an acute illness with moderate onset of symptoms (fever, malaise,
5 anorexia, nausea, abdominal discomfort, dark urine) and jaundice, and elevated
6 serum bilirubin and aminotransferases levels later on. The incubation period of
7 hepatitis A ranges from 15 to 50 days and clinical illness usually does not last
8 longer than 2 months, although 10–15% of patients have prolonged or relapsing
9 signs and symptoms for up to 6 months (Sjogren et al., 1987; Glikson et al., 1992).
10 In fact, with the advent of new highly sensitive techniques even in normal clinical
11 courses a high and long lasting viremia has been detected (Costafreda et al., 2006),
12 with the peak (up to 10^7 genome copies/ml of sera) occurring at 2 weeks after the
13 onset of symptoms and lasting up to an average of 6 weeks after the start of
14 symptoms (Bower et al., 2000; Costafreda et al., 2006). However, there is no evi-
15 dence of chronicity of the infection. Hepatitis A infection may occasionally pro-
16 duce fulminant hepatitis, mainly among patients with underlying chronic liver
17 diseases (Akriviadis and Redeker, 1989; Previsani et al., 2004).

The distribution patterns of hepatitis A in different geographical areas of the
18 world are closely related to their socioeconomic development (Gust, 1992; Hol-
19 llinger and Emerson, 2001; Previsani et al., 2004). The endemicity is low in devel-
20 oped regions and high in underdeveloped countries. The epidemiological pattern
21 has important implications on the average age of exposure and hence, as above
22 stated, on the severity of the clinical disease. Since hepatitis A infection induces a
23 life-long immunity (Hollinger and Emerson, 2001), severe infections among adults
24 are rare in highly endemic regions where most children are infected early in life. In
25 contrast, in low endemic areas the disease occurs mostly in adulthood, mainly as a
26 consequence of traveling to endemic regions, or as food or waterborne outbreaks,
27 and hence the likelihood of developing severe symptomatic illness is high. An
28 epidemiological shift, from intermediate to low prevalence, has been noticed in
29 recent decades in many countries, particularly in southern Europe, including Spain,
30 Italy and Greece (Germinario and Lopalco, 1999; Salleras, 1999; Van Damme et
31 al., 1999). Consequently, the Mediterranean basin as a whole should no longer be
32 considered as an endemic area (Previsani et al., 2004; Pintó et al., 2006). Addi-
33 tionally, some other countries from eastern Europe (Cinciara, 2000; Tallo et al.,
34 2003) have also described significant declines in the incidence of hepatitis A. AU :2

35 **General features of hepatitis A virus (HAV)**

36 The etiological agent of hepatitis A is the hepatitis A virus (HAV) which belongs to
37 genus *Hepatovirus* within family *Picornaviridae*, and as such it consists of a non-
38 enveloped icosahedral capsid of around 30 nm in diameter containing a positive
39 ssRNA genomic molecule of 7.5 Kb (Fauquet et al., 2005). The genome contains a
40 single open reading frame (ORF) encoding a polyprotein of around 2225 amino
41 acids (aa) preceded by a 5' non-coding region (5'NCR) that makes around 10% of
42

1 the total genome, and followed by a much shorter 3'NCR that contains a poly(A)
2 tract (Baroudy et al., 1985; Cohen et al., 1987). This genome is uncapped but
3 covalently linked to a small viral protein (VPg) (Weitz et al., 1986). The singly AU :3
4 translated polyprotein is subsequently cleaved into 11 proteins through a cascade of
5 proteolytic events brought about mainly by the viral 3C protease (Schultheiss et al.,
6 1994; Schultheiss et al., 1995). However, although the general genomic organiza-
7 tion and the expression pattern of HAV are very similar to those of most pico-
8 naviruses (Hollinger and Emerson, 2001; Agol, 2002), many differences exist
9 which deserve a special attention.

11 ***What makes hepatitis A virus such a special picornavirus***

12 The genetic distance between the genus *Hepatovirus* and the other genera of the
13 family reflects not merely a difference in the nucleotide and amino acid composition
14 but a difference in the molecular and biological characteristics of HAV. From the
15 genomic and proteomic points of view, several interrelated key issues must be
16 brought up. First of all, the structure of the 5'NCR and its internal ribosome entry
17 site (IRES). It is likely that picornavirus IRES has evolved by gradual addition of
18 domains and elements that improved its function in ribosome recruitment or oth-
19 erwise conferred regulation to the process of viral protein synthesis in a specific cell
20 environment (Ehrenfeld and Teterina, 2002). The HAV IRES is unique among
21 picornaviruses and constitutes the type III model (Braun et al., 1994; Ehrenfeld and AU :4
22 Teterina, 2002), which shows a very low efficiency in directing translation (Whetter
23 et al., 1994). Second, HAV encodes only a protease, 3C, while other picornavirus
24 code for additional proteases such as the L protease, in genus *Aphthovirus*, or the 2A
25 protease in *Enterovirus* and *Rhinovirus* genera (Leong et al., 2002). L and 2A
26 proteases, when present, play a crucial role in the primary cleavages of the viral
27 polyprotein while in those genera lacking these proteases, such as *Hepatovirus* and
28 *Paraechovirus*, both primary and secondary cleavages are conducted by the 3C
29 protease. But what is most important is that these additional proteases are involved
30 in the cellular protein shutoff induction (Leong et al., 2002). Since picornaviruses
31 utilize a mechanism of translation that is cap-independent and IRES-dependent,
32 the inhibition of non-essential cap-dependent cellular translation could be advan-
33 tageous to the virus. In doing so, the cellular translation machinery is utilized
34 almost exclusively for the production of viral proteins (Kuechler et al., 2002). An
35 early event preceding the shutoff of host cell protein synthesis is the cleavage of the
36 cellular translation initiation factor eIF4G, and evidence exists supporting that the
37 enzymes responsible of such a cleavage are 2A and L proteases in enteroviruses and
38 rhinoviruses, and aphthoviruses, respectively (Kuechler et al., 2002). An immediate
39 consequence of the lack of any of these proteolytic activities in HAV is its inca-
40 pacity to induce cellular shutoff which otherwise is directly related with its re-
41 quirement for an intact uncleaved eIF4G factor for the formation of the initiation
42 of translation complex (Borman et al., 1997; Jackson, 2002). An intriguing evo-
43 lutionary question remains to be solved regarding the selection of such an

1 inefficient IRES in HAV. An explanation has been suggested which accounts for
2 the constraints caused by the need to accommodate the dual functions of trans-
3 lation and replication in adjacent regions of the 5'NCR, generating inadvertent
4 consequences for either function (Ehrenfeld and Teterina, 2002). What has been
5 described up to now denotes that HAV must inefficiently compete for the cellular
6 translational machinery and thus it presents a unique translation strategy. This
7 points out to the third difference between HAV and other picornavirus members:
8 the codon usage. HAV presents a higher codon usage bias compared to other
9 members of its family, which conveys in the adaptation to use abundant and rare
10 codons (Sánchez et al., 2003b). In fact, 14 aa families contain rare codons, defined
11 in terms of their frequencies, making a total of 22 used rare codons. But what is
12 more surprising is that the HAV codon usage has evolved to be complementary to
13 that of human cells, never adopting as abundant codons those abundant for the
14 host cell, and even in some instances using these latter as rare codons. This dis-
15 parity, unique to HAV, has been interpreted as a subtle strategy to avoid, as much
16 as possible, competition for the cellular tRNAs in the absence of a precise mech-
17 anism of inducing shutoff of cellular protein synthesis (Sánchez et al., 2003b). As
18 stated before, a consequence of this special codon bias is an increase in the number
19 of rare codons used by HAV. Overall this increment is the result of the addition to
20 the cellular rare codons, also used as rare by the virus, of those most abundant
21 cellular codons that being unavailable for the virus are used at low frequencies.
22 Altogether, the HAV codon usage may contribute to its slow replication and to its
23 low yields. It has been largely documented, (Robinson et al., 1984; Sørensen et al.,
24 1989; Chou and Lakatos, 2004) the role of rare codons in the control of translation
25 speed, in the sense that clusters of rare codons would induce a transient stop of the
26 translational complex in order to seek for a suitable tRNA present at a very low
27 concentration among the pool of tRNAs. A function of these ribosome stallings
28 has been suggested to be the assurance of the proper folding of the nascent protein
29 (Adzhubei et al., 1996; Gavrillin et al., 2000; Evans et al., 2005). Such a function
30 may be postulated for HAV, where highly conserved clusters of rare codons strate-
31 gically located at the carboxi-ends of the structured elements have been reported
32 (Sánchez et al., 2003b). A certain contribution of the codon usage to the low
33 variability of the HAV capsid has been proposed taking into account that 15% of
34 its surface residues are encoded by such functional rare codons (Sánchez et al.,
35 2003b). This low capsid variability indeed correlates with a very low antigenic
36 variability: a single serotype exists, this being another striking difference with other
37 picornaviruses. The low capsid variability should rely on negative selection acting
38 against potential newly arising proteins, since the viral population replicates as a
39 quasispecies (Sánchez et al., 2003a). The quasispecies analysis revealed a dynamics
40 of mutation selection at and around the rare codons, confirming a seminal role of
41 the codon usage on HAV evolution (Sánchez et al., 2003a).

42 Other important differences exist between HAV and other picornaviruses at the
43 morphogenetic/structural level. The role of both ends (amino-VP4 and carboxi-2A)
of the capsid polyprotein in the virion assembly is still controversial (Probst et al.,

1 1999), and while there is no agreement on the requirement of VP4 for the mat-
2 uration of pentamers into capsids (Probst et al., 1999; Martin and Lemon, 2006), a
3 complete consensus exists on the necessity of 2A for pentamer formation (Probst et
4 al., 1999; Martin and Lemon, 2006). The ulterior removal of 2A in the mature
5 virion must be performed by a host cell protease (Graff et al., 1999; Martin et al.,
6 1999), although the mature 2A protein has never been identified directly in infected
7 cells. AU :6

8 The X-ray crystallographic structure has not yet been solved, due to the low
9 viral yields obtained by *in vitro* replication. However, recent 3D images of HAV
10 produced by cryoelectron microscopy (Holland Cheng, unpublished results) have
11 revealed important data being the most intriguing the lack of a well-defined canyon
12 around the fivefold axis of symmetry. The pit region of many picornaviruses con-
13 tains the receptor binding residues (Rieder and Wimmer, 2002) playing, thus, an
14 important biological role. A human HAV receptor (huhavr-1) has been identified
15 (Feigelstock et al., 1998), which contains both an amino terminal Ig-like domain
16 followed by a mucin-like domain (Silberstein et al., 2003). Huhavr-1 has been
17 detected in several human tissues, including the liver. Alternatively the asialoglyco-
18 protein receptor, to which IgA binds to, has been described to enable HAV in-
19 ternalization provided that the virus is complexed with such immunoglobulin
20 (Dotzauer et al., 2000). However, whichever is the receptor, the capsid region
21 involved in such an interaction remains to be elucidated. In contrast, the capsid
22 region interacting with the glycoporphin A of the human erythrocytes is indeed
23 located around the putative pit area (Sánchez et al., 2004a,b). The capsid structure,
24 however, is such that it tolerates this interaction only to occur at acid conditions,
25 being impaired at neutral biological conditions. Erythrocyte glycoproteins may
26 function as decoy receptors attracting pathogens to the erythrocyte and keeping
27 them away from target tissues (Gagneux and Varki, 1999), and hence the actual
28 capsid conformation that allows escaping from erythrocyte attachment may con-
29 stitute an advantage for a viremic infectious agent whose target organ is the liver.
30 In fact, pathogenesis is in part determined by the spread of the virus to the target
31 tissues (Rieder and Wimmer, 2002). In this context, key factors for the viral bi-
32 ological cycle and infection outcome are a high stability to the acid pH of the
33 stomach during the entry phase, a safe viremic phase, and resistance to the action
34 of detergents, particularly biliary salts, during the exit phase. This extreme resistant
35 phenotype of HAV explains its high persistence in the environment (Abad et al.,
36 1994a,b) and its transmission by contaminated foods and drinking water (Reid and
37 Robinson, 1987; Roseblum, 1990; Bosch et al., 1991; Dentinger et al., 2001;
38 Sánchez et al., 2002), which probably are the result of a highly cohesive capsid
39 conformation mediated through a very accurate folding.

41 *Hepatitis A transmission*

42 Hepatitis A is shed in the feces of infected patients. The viral concentration in such
43 stools is highest (up to 10^{11} genome copies/g of feces) after two weeks of the onset

1 of symptoms and lasts at least four more weeks (Costafreda et al., 2006). HAV
2 infection is mainly propagated via the fecal-oral route as the person-to-person
3 contact is the most common mode of transmission (Mast and Alter, 1993). In fact
4 HAV survival in contaminated fomites, such as sanitary paper, sanitary tile and
5 latex gloves, is very long (Abad et al., 1994a). In consequence, given the high
6 excretion level of HAV, transmission of the infection is facilitated when poor sani-
7 tary conditions occur. Nevertheless, transmission through the parental route may
8 occasionally occur (Noble et al., 1984; Sheretz et al., 2005).

AU:7

11 *Contaminated water and food as critical elements in the route of transmission*

13 Viruses present in the stool of infected patients are discharged into sewage which
14 ultimately may contaminate surface waters and seawater, and consequently be
15 acquired and concentrated by shellfish growing in these waters, or contaminate the
16 vegetables irrigated with the polluted waters. While in approximately 40% of the
17 reported cases of hepatitis A the source of infection cannot be identified, water-
18 borne and foodborne outbreaks of the disease have been reported. Within this
19 latter category, shellfish grown and harvested from waters receiving urban con-
20 taminants is a cause of large outbreaks of infectious hepatitis (Halliday et al., 1991;
21 Sánchez et al., 2002). Additionally, large outbreaks associated with the consump-
22 tion of berry fruits (Reid and Robinson, 1987) and vegetables (Roseblum et al.,
23 1990; Dentinger et al., 2001) irrigated with contaminated waters may occur. Wa-
24 terborne outbreaks are less common since the introduction of drinking water
25 treatments. However, reports exist when these measures fail and outbreaks of
26 hepatitis A occur (Bosch et al., 1991).

29 *Detection and quantification of hepatitis A virus*

31 As above stated, HAV may contaminate different types of environmental samples
32 including sewage, surface waters and seawater, and ultimately drinking water and
33 foodstuffs. Thanks to its high stability under very different conditions (Abad et al.,
34 1994b; Bosch et al., 1994), HAV can persist long enough in these edible samples to
35 be transmitted through their ingestion. Thus, although not compulsory, the screen-
36 ing of HAV presence is advisable in specific samples, at least when suspicion of
37 contamination exists.

39 Cell culture propagation of wild-type strains of HAV is a complex and tedious
40 task, which requires virus adaptation before its effective growth, and even in this
41 case, the virus usually establishes persistent infections resulting in low virus yields
42 (Flehmg, 1980; Daemer et al., 1981; Wang et al., 1986). Thus, infectivity is not
43 nowadays a useful method for primary HAV detection. Alternatively, immuno-
logical and, particularly, molecular techniques should be used.

1 *Molecular methods versus immunological methods: a clear choice in water virology*

3 Although fecal excretion of HAV is high, environmental samples usually contain
5 low viral numbers due mainly to the effect of dilution but also to some extent of
7 virus inactivation. Most of the immunological techniques used in clinical diagnosis
9 are aimed at the detection of those antibodies raised upon virus infection, mainly
11 IgM and IgG anti-HAV (Nainan et al., 2006), while antigen detection is uncom-
13 mon. Additionally, to the lack of immunological kits for antigen detection, their
15 sensitivity would not be high enough to be employed in scenarios of low environ-
17 mental virus concentration. All this calls for the development of highly sensitive
19 methods for HAV detection, such as those based on nucleic acids amplification.
21 The adoption of these techniques requires selection of the most adequate ampli-
23 fication target. The target region should be highly conserved to increase the chance
of detection and with an appropriate structure and length to allow the required
sensitivity. Immunological evidence determines the existence of a single serotype of
HAV (Lemon and Binn, 1983), although genomic analysis of the virus allows the
differentiation of six genotypes (Robertson et al., 1992; Costa-Mattioli et al., 2003).
However, all six genotypes are very closely related in the 5'NCR, which is the most
conserved region of the genome due to its functional structure in the processes of
translation and replication, as above stated, and with a maximum nucleotide di-
vergence of less than 5%. Consequently, the 5'NCR is the region of choice for the
design of broad spectrum molecular techniques.

25 *Genetic diversity*

27 In spite of the low antigenic variability of HAV, a degree of nucleotide variability
29 similar to that of other picornavirus in the capsid coding region has been described
(Sánchez et al., 2003b). This is explained by a very low number of non-synonymous
31 mutations per non-synonymous site and a number of synonymous mutations per
33 synonymous site similar to that of other picornaviruses. This genetic diversity
35 allows the differentiation of HAV into several genotypes and subgenotypes.
37 Different genomic regions have been used to differentiate the genotypes, including
the carboxi-terminus of VP3, the amino-terminus of VP1, the VP1 × 2A junction,
the region spanning the carboxi-end of VP1 till the amino-terminus of 2B (VP1/
P2B), and finally the entire VP1 region (see the review of Nainan et al., 2006).
However, partial genomic sequences never will guarantee the reliability of the
complete VP1/2A region. As a matter of fact, the identification of some HAV
antigenic variants affecting residues not included in the genotyping regions (Costa-
Mattioli et al., 2002a; Sánchez et al., 2002; Gabrieli et al., 2004) could have been
elusive in such circumstances. This is the reason why the use of long genomic
regions covering at least the entire VP1 including its 2A junction, has recently been
recommended (Costa-Mattioli et al., 2002a; European HAV Network, unpublished
results) for a more broad molecular typing of HAV. However, the VP1 × 2A junc-
tion is still the genomic region most in use worldwide (Robertson et al., 1992). In

1 this region, seven genotypes were initially defined, whose genetic distance was
2 >15% nucleotide variation. After refining this classification through the addition
3 of more sequences, only six genotypes exist at the present time (Costa-Mattioli et
4 al., 2002a; Lu et al., 2004). Three out of these six genotypes (I, II and III) are of
5 human origin while the other (IV, V and VI) are of simian origin. Genotypes I and
6 II contain subgenotypes (Ia, Ib, IIa and IIb) defined by a nucleotide divergence of
7 7–7.5%.

8 Whether the objective is the general broad spectrum detection of HAV or the
9 typing of the isolated strains will determine the use of 5'NCR or VP1 × 2A targets
10 (Sánchez et al., 2004a,b) or, alternatively, other genotype targets.

11 *The advent of real-time standardized quantitative techniques for the accurate*
12 *estimation of the HAV titer*

13 Many molecular techniques have been used throughout the years for HAV detec-
14 tion, including hybridization (Bosch et al., 1991; Zhou et al., 1991) and amplifi-
15 cation techniques, essentially RT-PCR, usually combined with confirmation tests
16 such as the southern blotting (Calder et al., 2003; Sánchez et al., 2004a,b), re-
17 striction fragment length polymorphism (Goswami et al., 1997) or sequencing
18 (Sánchez et al., 2002). Recently, new approaches have been developed which enable
19 not only the detection but the quantification of the genome copy numbers in a real-
20 time scale. These techniques basically include two different alternatives: the real-
21 time RT-PCR and the nucleic acid sequence based amplification (NASBA). Both
22 techniques take advantage of the use of a combination of enzymes for the am-
23 plification of RNA and a reliable method to quantify the final amplified product,
24 usually based on the use of fluorescent probes. In the real-time RT-PCR, the gold
25 standard is the combination of the reverse transcriptase (RT) and the *Taq* polym-
26 erase to transform the target ssRNA into the final dsDNA, while the NASBA is
27 again the RT this time with the T7 RNA polymerase and a primer including the T7
28 promoter that allows the synthesis of great amounts of the final RNA product.
29 Both combinations show a high intrinsic amplification power and although it has
30 been claimed that the NASBA technique allows an even higher amplification than
31 the real-time RT-PCR, more descriptions exist on the use of this latter technique
32 (Costa-Mattioli et al., 2002b; Jothikumar et al., 2005; Costafreda et al., 2006) than
33 with the NASBA (Jean et al., 2001; Abd El Galil et al., 2005) for the quantification
34 of HAV.

35 Two key issues have to be solved when real-time quantification techniques are
36 developed. Finding those optimal conditions for each particular target rather than
37 using universal settings. It has been claimed that the use of one-step RT-PCR
38 formats reduces the HAV detection sensitivity by up to 1 log unit (Nainan et al.,
39 2006) in comparison to two-step formats. However, when optimum conditions are
40 established for each format, the same level of sensitivity is achieved irrespective of
41 the use of one-step or two-step formats (Costafreda et al., 2006). Not only is the
42 establishment of optimal conditions crucial in the success of a highly sensitive
43

1 quantification technique, but also standardization of the procedures. Detection of
2 viral RNA by molecular amplification procedures involves several essential steps,
3 the viral RNA extraction and the reverse transcription being the most critical ones.
4 Since no universal reliable nucleic acid extraction method exists, the best option is
5 to control the efficiency of extraction for a particular matrix. The addition of a
6 known concentration of an external virus control to the samples to be quantified
7 has been proposed to assess the efficiency of RNA extraction of enteric viruses in
8 different matrices. Vaccinal poliovirus type 2 (Nishida et al., 2003) and vaccinal
9 Mengo virus (Costafreda et al., 2006) have been proposed as such controls. How-
10 ever, poliovirus presents the inconvenience of being itself a potential contaminant
11 of samples and additionally its use in the context of the poliomyelitis eradication
12 era (see chapter by Hovi et al.) is not advisable. The control of the reverse tran-
13 scription reaction is more widely performed and most of the real-time techniques
14 described for the quantification of HAV include an RNA standard internal control
15 (Costa-Mattioli et al., 2002b; Jothikumar et al., 2005; Costafreda et al., 2006).
16 Usually, these controls are synthesized by *in vitro* transcription of cloned cDNAs or
17 T7 promoter-containing amplimers corresponding exactly to the targets, and are
18 added to each test tube at known amounts in order to follow up the efficiency of the
19 reaction. Applying these control measures, a fine estimation of the number of HAV
20 genome copies per gram of shellfish was performed for the first time in several clam
21 stocks associated with an outbreak of hepatitis A, and ranged from 1×10^3 to
22 1×10^5 (Costafreda et al., 2006). These determinations have provided the means for
23 a risk-assessment study linking the level of shellfish contamination and the attack
24 rate of infection (Costafreda et al., unpublished results).

25 *Molecular epidemiology of hepatitis A: the water environment* 26 *as an overview of the circulating viruses*

27 Environmental surveillance, mostly sewage monitoring, has been used to assess the
28 circulation of different human enteric viruses, notably rotaviruses (Villena et al.,
29 2003; van Zyl et al., 2006), wild-type poliovirus (Deshpande et al., 2003; El Bassioni
30 et al., 2003) and also HAV (Pintó et al., 2006) as well as animal enteric viruses
31 (Jiménez-Clavero et al., 2003, 2005a). The application of molecular typing tech-
32 niques in this kind of samples allows the molecular and phylogenetic analysis
33 required for the traceability of strains, i.e. in the context of the poliovirus erad-
34 ication program (see chapter by Hovi et al.). The molecular characterization of
35 viral strains isolated in sewage, i.e. rotavirus, might be very useful in the policy for
36 the design for future vaccines applicable to countries where the clinical surveillance
37 does not cover all the population.

38 Environmental molecular epidemiology as such should rely not only on the
39 molecular characterization of the isolated strains but also on the epidemiological
40 data regularly collected. When both disciplines are applied together interesting
41 conclusions might be drawn. In the particular case of HAV, a study relating a
42 vaccination campaign, the number of clinical cases and the occurrence of viruses in
43

1 sewage during a period of 5 years in Barcelona city has been reported (Pintó et al.,
2006). Attack rates per 100,000 inhabitants of 9.1, 6.2, 3.3, 1.7 and 8.0 were es-
3 timated during the years 1998–2002. While the progressive decline could be clearly
associated to a vaccine administration the final increase was attributed to the huge
5 immigration flow, from North Africa, South America and East Asia that the city
received (nearly 10% of the total population). The vaccination campaign was, and
7 is, dedicated to children 12 years old, being those <12 still susceptible to the
infection. Immigrant children may act as potential carriers of the infection, and in
9 fact many of the observed cases were school-related, and mostly coincided with the
virus incubation period elapsed after the return from the school holiday, and after
11 the immigrant population have returned from visits to their countries of origin.
Sewage surveillance data taught us that the similar high attack rates seen at the
13 beginning and final years of the study responded to a different infection pattern.
While many asymptomatic cases might occur at the beginning with high levels of
15 virus excreted and with many positive isolations in sewage, at the end only small
outbreaks among the non-vaccinated population arose without the massive in-
17 volvement of asymptomatic carriers and consequently without positivity in sewage.
In conclusion, the environmental study reflected that in spite of the increase in
19 clinical cases in the last year of study, the vaccination program was anyway work-
ing well.

21 Finally, it should be stated that molecular environmental surveillance still
presents some intrinsic problems, genotyping being one of them. Even when using a
23 short fragment such as the VP1 \times 2A region, HAV genotyping is a hard task due to
the low concentration of viruses occurring in sewage. Using this genotyping target,
25 a threshold of 10^5 genomes/ml has been estimated. In consequence, only those
genotypes, more prevalent in the population, are likely to be detected. However in
27 areas endemic for HAV, such as Egypt (Pintó et al., 2006), environmental surveil-
lance is a powerful tool to complement the clinical epidemiology data, since de-
29 tection and genotyping as well are possible. In fact, as above stated, this approach
has been already implemented in Egypt in the context of the polio eradication
31 program (El Bassioni et al., 2003).

33 *Prevention of hepatitis A*

35 Inactivated HAV vaccines are available since the early 1990s and provide long-
lasting immunity against hepatitis A infection (Bell and Feinstone, 2004). The
37 immunity is largely related to the induction of high titers of specific antibodies.
Thanks to the existence of a single serotype of HAV, these vaccines are of high
39 efficacy. These vaccines consist of viruses grown in cell culture, purified, inactivated
with formalin and adsorbed to an aluminum hydroxide adjuvant, making their
41 economic cost quite high. This is the reason why many discrepancies already exist
on their universal use in massive vaccination campaigns. Countries with previous
43 intermediate endemicity of HAV such as Israel or some autonomous communities
of Spain such as Catalonia, or some States of United States have performed studies

1 on the impact of child vaccination on the overall incidence of hepatitis A con-
2 cluding that the immunization is medically (Salleras, 1999; Shouval, 1999; Wasley
3 et al., 2005) and economically (Dagan et al., 2005; [http://www.gencat.net/salut/](http://www.gencat.net/salut/portal/cat/nen.htm)
4 <http://www.cdc.gov/nip/publications/VIS/vis-hep-a.pdf>)
5 justified. In contrast, other countries in a similar situation such as Italy do not
6 recommend at present the implementation of such a measure in terms of cost-
7 benefits (Franco and Vitiello, 2003). In this context is quite evident that high
8 endemic countries that usually have low economic incomes do not regard the
9 vaccination against hepatitis A as a primary policy (Teppakdee et al., 2002).

10 Although several attenuated vaccine candidates have also been attempted, due
11 to the successful use of inactivated vaccines, its development is hardly plausible.

12 As a general rule, in low and intermediate endemic regions, where paradoxically
13 the severity of the disease is high, vaccination against hepatitis A should be rec-
14 ommended in high-risk groups, including travelers to high endemic areas, men
15 having sex with men, drug users and patients receiving blood products. In addition,
16 the inclusion of hepatitis A vaccines in mass vaccination programs in those coun-
17 tries receiving high numbers of immigrants from endemic countries is particularly
18 advisable. However, and bearing in mind the quasispecies replication pattern of
19 HAV (Sánchez et al., 2003a) that could lead in populations with continued ex-
20 posure to the virus to the selection of new antigenic variants escaping immune
21 protection, mass vaccination programs in highly endemic areas is controversial.

22 **Hepatitis E**

23 *Natural course and epidemiology of hepatitis E*

24 Hepatitis E, previously known as enterically transmitted non-A, non-B hepatitis, is
25 an infection with clinical and epidemiological features of acute hepatitis. The clin-
26 ical presentation of hepatitis E is basically similar to that of hepatitis A, but
27 cholestatic jaundice is more common. The clinical course of the infection was first
28 addressed in a human volunteer that ingested a clarified stool preparation from an
29 infected patient (Balayan et al., 1983). The incubation period averages 40 days,
30 with a range between 15 and 60 days. Amino alanine transferase elevation occurs
31 during 30–120 days after infection, and fecal excretion of the virus begins around 1
32 week before onset of illness and continues for, at least, 2–3 weeks thereafter (Skid-
33 more, 2002). The ecteric phase of the infection is characterized by a flu-like prod-
34 rome with epigastric pain, vomiting, fever and discoloration of the urine, but
35 jaundice patients display yellowish skin, scleral icterus, dark urine, and light tan-
36 colored feces; however most infections are asymptomatic and no evidence of
37 chronic disease has been observed (Purcell and Emerson, 2001; Smith, 2001). As a
38 result of viral replication in the liver, the hepatitis E virus (HEV) is found in the bile
39 in large quantities, reaching the intestines by the bile duct and being subsequently
40 shed in the feces. Asymptomatic infected individuals may shed virus and become
41 reservoirs of the virus between epidemics and, therefore, contribute to sporadic
42

1 infection by person-to-person transmission or by contaminating water and food. In
2 endemic regions, the overall attack rate was estimated to be around 2.5% in adults
3 and around 1.2% in children (Vishwanathan, 1957). The case-fatality rate is usually
4 low (0.2–3%), but in pregnant women during the third trimester of gestation it can
5 be as high as 15–25%, primarily due to fulminant hepatic failure (Purcell and
6 Emerson, 2001; Smith, 2001; Skidmore, 2002).

7 The significance of the host immune response in the pathogenesis of cell damage
8 is not fully understood. Anti-HEV IgM appears at the time of the onset of
9 symptoms and remains detectable for to 2–3 months. Anti-HEV IgG is detectable
10 shortly after IgM detection, increases during the acute phase and may be present in
11 serum for years after the initial infection (Clayson et al., 1995b).

12 There is no specific treatment for hepatitis E. Passive immunization with
13 convalescent sera has been accomplished in animal models (Tsarev et al., 1994) but in
14 humans administration of immune globulin obtained from inhabitants of HEV
15 endemic regions was unsuccessful (Khuroo and Dar, 1992); however, it should be
16 noted that this study used unselected plasma and that, even in endemic regions,
17 anti-HEV prevalence and titers are low. Therefore, it cannot be ruled out that the
18 use of selected anti-HEV batches of immunoglobulins with high titers may be
19 useful, particularly for pregnant women and/or during epidemics.

20 HEV is transmitted primarily by contaminated water, and causes frequent
21 epidemics in areas with inadequate water supplies and poor sanitary conditions
22 (Purcell and Emerson, 2001), being the principal cause of acute, sporadic hepatitis
23 in adults in many areas of Asia, Middle East and Northern Africa (Emerson and
24 Purcell, 2003; Schlauder, 2004). Recently, an increase in the number of cases in
25 regions considered as non-endemic for hepatitis E has been reported (Smith, 2001;
26 Worm et al., 2002a).

27 Since the main route of HEV transmission is feco-oral, most epidemics can be
28 linked to waterborne outbreaks, particularly in developing countries with warm
29 weather, high population density and poor sanitary conditions. The first documented
30 outbreak of HEV occurred in India in 1955–1956 (Vishwanathan, 1957).
31 The origin of the outbreak, which was initially attributed to hepatitis A and later
32 on confirmed to be hepatitis E, was the contamination by sewage, from 1 to 6 weeks
33 prior to the epidemic, of Jumna River, the source of water for the treatment plant.
34 Alum and chlorine treatment prevented bacterial infections, but 30,000 cases of
35 hepatitis occurred among the population (Wong et al., 1980). One of the highest
36 epidemic areas is China, where at least 11 epidemic outbreaks have been reported to
37 date. The largest one occurred in 1986–1988, with more than 119,000 cases that
38 resulted in more than 700 deaths. Until recently most cases reported in developed
39 countries were attributed to travel to endemic areas, however, as aforementioned,
40 there is an increase in the number of cases of infected patients that had never been
41 abroad (Smith, 2001; Worm et al., 2002a).

42 In addition, a zoonotic potential for the virus was suggested after detection of
43 HEV infection in wild and domestic animals, and later on confirmed in people who
44 ate HEV infected uncooked deer meat (Meng, 2003; Goens and Purdue, 2004). The

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1 risk of a zoonotic spread of the virus, its detection in non-endemic areas, and the
2 continuous occurrence of outbreaks in endemic regions have boosted the interest in
3 the understanding of the biology and life cycle of the virus, and in the improvement
4 of diagnostic tools able to detect the pathogen in polluted waters and other en-
5 vironmental samples.

7 *Seroprevalence*

9 The overall anti-HEV prevalence reported in endemic countries is quite variable,
10 but lower than expected, 3–27% (Purcell 1994; Purcell and Emerson, 2001; Worm
11 et al., 2002b). In contrast to other enteric viruses such as poliovirus or HAV, the
12 prevalence of anti-HEV IgG is lower in children than in adults (Arankalle et al.,
13 1995, Meng, 2002). A possible explanation for this could be that HEV immunity
14 acquired with subclinical infection during childhood wanes with time. In non-
15 endemic areas with good sanitary conditions and control of water supplies, there is
16 a low but constant increase in the number of HEV sporadic cases non-related to
17 travel, and the anti-HEV antibody prevalence among the healthy population is
18 relatively high, even higher than that reported in endemic areas (Meng et al., 2002;
19 Worm et al., 2002a; Meng et al., 2003).

20 After the first description of HEV in swine (Balayan et al., 1990; Clayson et al.,
21 1995a; Meng et al., 1997), an initial epidemiological survey in North America
22 reported a higher prevalence of anti-HEV antibodies among swine veterinarians
23 (26%) than among blood donors (18%) (Meng et al., 2002). A further study de-
24 scribed a 35, 11 and 2.5% seroprevalence among swine, swine workers and non-
25 swine workers, respectively (Whiters et al., 2002). Recently, a Chinese study has
26 reported that swine workers have a 74% higher risk of HEV infection than people
27 engaged in other occupations (Zheng et al., 2006). In contrast, no statistical differ-
28 ence in anti-HEV antibodies prevalence has been noted between pig farmers
29 (13.0%) and control subjects (9.3%) in Sweden (Olsen et al., 2006). In any case, it
30 should be noted that differences in study design (population features, health status,
31 demographical variables, etc...) make difficult the comparison of the reported
32 data, and that there are contradictory results about the reliability of the anti-HEV
33 detection test used in the different studies (Worm et al., 2002a; Emerson and
34 Purcell, 2003; Schlauder, 2004). Hence, analyses of well-selected population with
35 standardized reagents are needed to have a more clear understanding of the actual
36 incidence of HEV infection.

37 *General features of the hepatitis E virus (HEV) with a special emphasis* 38 *to the genomic organization*

41 The identity of the causative agent of hepatitis E was first described in 1990 (Reyes
42 et al., 1990). One year latter, the entire sequence of the viral genome was published
43 (Tam et al., 1991). HEV was provisionally classified as a member of the *Caliciviridae*
family, but it is now ascribed to a separate family, *Hepeviridae*, in the

1 prototypic genus *Hepevirus* (Mayo and Ball, 2006). HEV is a spherical, non-en-
2 veloped viral particle of around 32–34 nm in diameter. The genome is a ssRNA
3 molecule of positive polarity of approximately 7.2 Kb containing 3 overlapping
4 ORF and a 3'poly (A) tail (Worm et al., 2002b; Emerson and Purcell, 2003; **AU:12**
5 Schlauder, 2004). **AU:13**

In vitro analysis suggested that HEV RNA is capped at the 5'end (Kabrane-
6 Lazizi et al., 1999a). After a non-coding region of 27–35 nucleotides (nt), ORF-1
7 encodes about 1693 aa encompassing non-structural proteins with enzymatic ac-
8 tivity that are involved in viral replication, transcription and protein processing,
9 including the viral replicase (Emerson and Purcell, 2003). ORF-2 extends 1980 nt,
10 terminating 65 nt upstream of the poly-A tail, and renders a 660 aa protein likely
11 representing the structural capsid protein(s) (Tam et al., 1991). *In vitro* experiments
12 suggested that ORF-2 protein is synthesized as a large glycoprotein precursor of
13 around 88 kDa, which is cleaved into the mature protein (Jameel et al., 1996,
14 Zafrullah et al., 1999). ORF-2 protein contains epitopes that induce neutralizing
15 antibodies and are mainly located near the carboxi-end (Tam et al., 1991). ORF-3
16 overlaps the 5'end of ORF-1 by only 1 nt and ORF-2 by 328 nt. It encodes a 123 aa
17 protein which is post-translationally modified by phosphorylation giving a mature
18 protein of around 13.5 kDa of unknown function (Emerson and Purcell, 2003).
19 This phosphoprotein is associated with the hepatocellular cytoskeleton (Zafrullah
20 et al., 1997) and form a complex with capsid protein of ORF-2 and, thus, it is
21 believed to be involved in the assembly of the viral particle (Jameel et al., 1996).
22 However, it has been recently shown that, in contrast to its requirement *in vivo*,
23 ORF-3 protein is not required for infection of Huh-7 cells or production of in-
24 fectious virus *in vitro* (Emerson et al., 2006). ORF-3 may also have regulatory
25 functions implicated in modulation of cell signaling (Emerson and Purcell, 2003).
26 In addition, ORF-3 protein also beards neutralizing epitopes near its 3'end (Tam
27 et al., 1991). In any case, it should be noted that the lack of a suitable and efficient cell
28 culture system for replication of HEV has hampered the study of the viral life cycle
29 (Emerson and Purcell, 2003).

30 *Genetic variation*

31
32 The genome sequence of HEV is quite stable (Arankalle et al., 1999). A high
33 genomic homology is found among isolates from the same outbreak, and serial
34 passages in animal models did not result in genetic drift (Worm et al., 2002b;
35 Schlauder, 2004). However, data supporting a quasispecies organization of HEV
36 genome during epidemics have also been reported (Grandadam et al., 2004). Ad-
37 ditionally, isolates from different geographical regions are relatively diverse. Based
38 on this genomic heterogeneity, HEV has been classified into four different gen-
39 otypes (Worm et al., 2002b; Meng, 2003; Schlauder, 2004). Genotype I is mainly
40 presented in endemic areas from Asia and Africa. Genotype II includes the Mex-
41 ican isolates and some Nigerian strains. Isolates from regions considered as non-
42 endemics (USA, Spain, Italy, Greece, etc.) represent a more diverse cluster of
43

1 sequences and are grouped into genotype III. Finally, genotype IV includes isolates
3 from China. Besides this genotypic diversity, no evidence of serological heteroge-
neity has been reported and, therefore, it seems that there is only one HEV se-
rotype.

5 ***HEV transmission***

7 *Waterborne transmission*

9 Epidemics of hepatitis E in endemic areas are usually due to fecally contaminated
11 water (Aggarwal and Naik, 1994) and most outbreaks can be traced back to con-
taminated water sources (Smith, 2001). Adequate circumstances for HEV epidem-
13 ics arise when raw sewage enters in contact with water reservoirs during heavy rain
sessions, floods, monsoons, etc. For instance, heavy and flooding rains preceded
15 the Indian epidemic of the 1950s (Vishwanathan, 1957; Khuroo and Kamili, 1994).
In most instances, people affected by HEV outbreaks lives near rivers with inad-
17 equate sanitary conditions (Bile et al., 1994) and a high incidence of HEV se-
ropositivity has been correlated with the use of non-boiled river water for drinking,
19 cooking and washing. Likewise, refugees and people living in urban crowded slums
and camps are at increased risk for fecal-oral transmitted diseases, including HEV
21 infection (Khuroo and Kamili, 1994; Mast et al., 1994), as it has been recently
demonstrated in the displaced population from Darfur (Sudan) where, in 6 months,
23 2.621 hepatitis E cases were recorded (attack rate 3.3%). The case-fatality rate was
1.7%, with 45 deaths, including 19 pregnant women (Guthmann et al., 2006).

25 HEV was detected in all sewage influent samples and in 67–89% of effluent
samples from sewage treatment plants in Madras, India, showing that treatment
27 was not as effective as it should be. Viral particles have also been detected in sewage
from industrialized countries (Pina et al., 2000). HEV RNA was detected in a
29 pretreated sewage sample collected in Washington, DC, and it showed a very high
homology with human and swine isolates from the US (Clemente-Casares et al.,
31 2003). In contrast, no HEV RNA has been detected in drinking and surface waters
collected from pig farms where HEV was present (Kasorndorkbua et al., 2005).

33 *Person-to-person transmission*

35 Person-to-person transmission seems to be low (Aggarwal and Naik, 1994; Bile et
37 al., 1994; Mast et al., 1994). Secondary cases among household members of patients
with documented HEV infection occurs in 1–2% (Aggarwal and Naik, 1994). Per-
39 son-to-person transmission in hospital settings has been described, although results
about the incidence of HEV infection in hospitalized patients and in people that
41 received contaminated blood are inconclusive (Smith et al., 2001). Data on mother-
to-child transmission rates of HEV are quite variable, ranging between 30 and
43 100% (Khuroo et al., 1995; Kumar et al., 2004). Additionally, it has been reported
that up to 2/3rd of pregnant HEV-infected women may have preterm delivery

1 (Kumar et al., 2004). HEV RNA has also been detected in the blood of newborns at
2 a time when no virus was detectable in the mother (Khuroo et al., 1995).

3 *Foodborne transmission*

5 Washing, irrigating and processing of food with HEV-contaminated water could
6 lead to HEV outbreaks if the food is eaten uncooked. Food manipulation by an
7 HEV-infected person may also transmit the disease. Acute hepatitis E in Sicily
8 (Italy) was attributed to contaminated shellfish consumption (Cacopardo et al.,
9 1997), and a case of hepatitis E after ingestion of Chinese medicinal herbs has also
10 been reported (Ishikawa et al., 1995). Likewise, sporadic acute or fulminant hep-
11 atitis has been linked to uncooked pig liver and wild boar meat consumption in
12 Japan (Yazaki et al., 2003; Li et al., 2005). Finally, a clear demonstration of acute
13 HEV infection after consumption of HEV-infected uncooked deer meat has been
14 reported (Tei et al., 2003). In this latter study, 4 out of 5 individuals who ate the
15 infected meat presented hepatitis, while the 3 other members of the families who did
16 not eat it were not infected. One child who was not infected claimed to have eaten a
17 very small portion of deer meat, suggesting that HEV infection is dose-dependant.
18 Sequence analysis of HEV RNA from patients and from frozen leftover deer meat
19 showed a 100% similarity (Tei et al., 2003). AU:15

21 *Zoonotical transmission*

23 Presence of anti-HEV antibodies in pigs and characterization of swine HEV were
24 first described in the 1990s (Balayan et al., 1990; Clayson et al., 1995a; Meng et al.,
25 1997). Later on, experimental infection of pigs with either swine HEV or human
26 isolates was achieved, and showed that infected animals presented viremia and shed
27 virus in feces, although no clinical or biochemical signs of disease were observed
28 (Balayan et al., 1990; Meng et al., 1998; Halbur et al., 2001). After that, several
29 evidences have raised the hypothesis of a zoonotic potential for HEV and its possi-
30 ble risk in xenotransplantation (Meng, 2003). For instance, HEV has been de-
31 tected in sewage polluted with pig feces (Pina et al., 2000), people drinking water
32 from downstream of pig farms seem to have a higher risk of HEV infection (Zheng
33 et al., 2006) as do workers engaged in occupations related to swine farming (Meng,
34 2003; Zheng et al., 2006), and anti-HEV antibodies have been found in swine herds
35 from endemic and non-endemic areas (Emerson and Purcell, 2003; Meng, 2003;
36 Goens and Purdue, 2004). Furthermore, in general, swine isolates are genetically
37 more closely related to human HEV strains of the same geographical region than to
38 swine strains of other parts of the world (Meng et al., 1997; Meng, 2002; Meng et
39 al., 2003). Finally, as aforementioned, HEV infection in humans after ingestion of
40 HEV-infected raw deer meat has been demonstrated (Tei et al., 2003).

41 Besides pigs, specific anti-HEV antibodies and HEV strains have also been
42 detected in rodents (Clayson et al., 1995a; Kabrane-Lazizi et al., 1999b), wild boar
43 (Matsuda et al., 2003), donkeys (Guthmann et al., 2006), chickens, cattle and dogs

1 (Meng, 2003; Goens and Purdue, 2004). More recently, an avian HEV has been
2 described (Haqshenas et al., 2001) and, although it is genetically less related to
3 human HEV than swine isolates, it shares antigenic epitopes with both of them
4 (Haqshenas et al., 2002). All these observations have strengthened the zoonotic
5 potential of HEV, but the assessment of a zoonotic transmission to humans
6 through animal waste still needs further evaluation.

7 ***Diagnosis***

8 *Serological diagnosis*

9
10
11 Enzyme-linked immunosorbent assay (EIA) is the main diagnostic tool to detect
12 anti-HEV IgG and IgM (Worm et al., 2002b). In general, a positive result for anti-
13 HEV IgM indicates acute disease, however, to avoid false negative results, testing
14 should be done in the acute phase of the infection. Although detection of anti-HEV
15 IgG is not conclusive of HEV infection, a high IgG titer or increasing titers in
16 consecutive samples, support the diagnosis of acute hepatitis E.

17 Several antigenic domains have been identified in the three ORFs of HEV.
18 Based on this information, different synthetic peptides and recombinant proteins
19 derived from the carboxi-end of ORF-2 and/or ORF-3 have been assayed for
20 specific antibody detection (Worm et al., 2002a). Studies carried out with these tests
21 have reported a relatively high seroprevalence in non-endemic countries, raising
22 concerns about the possible detection of non-specific cross-reactive antibodies
23 (Mast et al., 1998; Worm et al., 2002a). However, a blind comparison of a test
24 based on a recombinant ORF-2 protein showed that it was 98% specific for anti-
25 HEV (Mast et al., 1998). When this approach was applied to sera from different
26 regions of the world, it confirmed the previous results obtained with other assays
27 (Emerson and Purcell, 2003). At present, the few commercially available EIAs kits
28 are based on the Mexican and/or the Burmese prototypes, and although a recent
29 comparison of different tests using outbreak samples has shown that they can be
30 highly specific and sensitive (Myint et al., 2006), their reliability still needs to be
31 fully confirmed worldwide.

32 *Molecular detection*

33
34
35 Detection of HEV by RT-PCR is indicative of active infection. The availability of
36 an increased number of HEV sequences from different sources and geographical
37 regions has enabled the design of specific oligonucleotide primers that match con-
38 served regions of the HEV genome and allows the detection of HEV in acute phase
39 sera, stools and contaminated water and sewage (Schlauder et al., 2004). Several
40 conventional “in-house” RT-PCR assays have been published for detection of
41 HEV in serum, feces and bile of infected individuals (Smith, 2001). For water
42 analysis, efficient concentration procedures and highly sensitive detection methods
43 are required for viral detection (Jiménez-Clavero et al., 2005b; and chapter by

1 Wyn-Jones). Recently, several real-time RT-PCR detection methods have been
2 described (Mansuy et al., 2004; Orrù et al., 2004; Enouf et al., 2006; Jothikumar et al., 2006). Using an internal control and spiked water samples, detection of as few
3 as 4 genome equivalent copies of HEV plasmid DNA and of 0.12 pig ID₅₀ of swine
4 HEV has been achieved (Jothikumar et al., 2006). Development of a quantitative,
5 broadly reactive, quick, easy and reproducible HEV detection method would be of
6 special interest for testing water and environmental samples, and may allow track-
7 ing of the polluting sources. Recently, as few as 100 fM of an ORF-2 amplicon were
8 detected using an HEV specific microarray (Liu et al., 2006). Ideally, development
9 of a microarray assay able to detect as much waterborne pathogens as possible in a
10 single reaction would greatly improved our current capacity for detection of water
11 pollutants representing human and/or animal health risk.

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15 *Prevention of hepatitis E*

16
17 The feasibility of HEV vaccines is based on several evidences: (i) specific antibodies
18 are raised after HEV infection; (ii) HEV infected people are usually protected
19 following epidemics; and (iii) animal experimentation has shown that passive im-
20 mune prophylaxis induces humoral immunity (Emerson and Purcell, 2001; Wang
21 and Zhuang, 2004). Additionally, only one HEV serotype has been described, thus,
22 production of a broadly cross-reactive vaccine should be possible. Such a vaccine
23 would be useful in protection against HEV infection, mainly in pregnant women
24 and in people from endemic regions and travelers to these areas. Nevertheless, the
25 lack of a susceptible cell culture system has hampered the development of live
26 attenuated or killed vaccines (Wang and Zhuang, 2004) and hence no commercial
27 vaccines against HEV are available.

28 To date, most research on HEV vaccines is focused on ORF-2-derived proteins
29 or peptides that contain neutralizing epitopes common to different genotypes
30 (Emerson and Purcell, 2001; Meng, 2001; Worm et al., 2002a; Wang and Zhuang,
31 2004). Several ORF-2 vaccine candidate products have been expressed in insect,
32 prokaryotic, yeast, animal and plant cells (Emerson and Purcell, 2001; Wang and
33 Zhuang, 2004). Animal experimentation has shown that administration of some of
34 these ORF-2 recombinant proteins protected against homologous and het-
35 erologous challenge (Purdy et al., 1993; Tsarev et al., 1994; Tsarev et al., 1997;
36 Im et al., 2001). DNA immunization of mice with an HEV-cDNA elicits high titers
37 of specific anti-HEV antibodies (He et al., 1997), immunologic memory (He et al.,
38 2001) and protection in cynomolgus macaques (Kamili et al., 2004). Truncated
39 ORF-2 protein expressed in baculovirus spontaneously assembles into viral-like
40 particles (VLPs) and are also good immunogens (Li et al., 2001). A recombinant
41 HEV baculovirus vaccine candidate that protects against intravenous administra-
42 tion of heterologous HEV strains has entered into preclinical trials (Stevenson,
43 2000; Zhang et al., 2002; Purcell et al., 2003).

1 *Concluding remarks*

3 Enterically transmitted hepatitis represent by large the most common manifestation
5 of acute hepatitis worldwide. Regarding hepatitis A, although the increase in living
7 standards and public health sanitation are greatly contributing to a decrease in its
9 global incidence, the total number of cases per year is still extremely high with
11 estimations up to the scale of millions. Overall the situation encourages policies of
immunization, but in spite of the availability of vaccines, their use in highly en-
demic regions may be limited on the basis of their cost and even on the basis of
medical and molecular aspects. Thus in these areas prevention should, at least, rely
on the implementation of effective control measures such as water sanitation and
virus monitoring.

13 In the case of hepatitis E, the disease that was restricted to endemic areas is now
15 been increasingly reported also in regions considered as non-endemic. However, it
17 remains unclear whether this increase is related to the emergence of the pathogen or
19 to the new epidemiological and public health interest in this virus and the avail-
21 ability of diagnostic procedures. This concern derives from the associated mortality
in pregnant women, as well as the morbidity and disability in the general pop-
ulation, mainly in endemic areas. Since HAV and HEV are transmitted by con-
taminated waters, improvement of water quality by proper sewage disposal and
water treatment is very important in preventing spread of these infections.

23 Although it may appear unlikely that infectious hepatitis outbreaks occur in
25 industrialized countries with properly treated waters and sewages and good sani-
27 tation and hygienic conditions, potential risk derived from consumption of veg-
etables, fruits and other products imported from endemic regions exist. Bivalve
mollusks grown and harvested in polluted waters represent a particular potential
threat due to their capacity to filter large volumes of water and accumulate the
viruses in their edible tissues.

29 The possible zoonotic transmission of HEV may also contribute to the con-
31 tinuous spread of the virus, despite improved sanitation, and calls for a more deep
33 knowledge of HEV prevalence in animals in order to prevent hepatitis E trans-
mission.

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37 Martin et al.; Meng et al., 2001.

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