i-motif structures in long cytosine-rich sequences found upstream of the promoter region of the SMARCA4 gene
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1.Overview

- Cytosine-rich oligonucleotides are capable of forming complex structures known as i-motif with increasingly studied biological properties [1,2].
- The study of sequences prone to form i-motifs located near the promoter region of genes may be difficult because these sequences not only contain repeats of cytosine tracts of disparate length but also these may be separated by loops of varied nature and length [3].
- In this work, the formation of an intramolecular i-motif structures by a long sequence smcon located upstream of the promoter region of the SMARCA4 gene has been demonstrated. NMR, CD, Gel Electrophoresis, SEC, and multivariate analysis have been used. Not only the wild sequence (5'-TC₃T₂GCTATC₃TGTC₂TGC₂TCGC₃ T₂G₂TCATGA₂C₄-3') has been studied but also several other truncated and mutated sequences.

2. NMR data



¹H NMR spectra of SMC01 (a), SMC02 (b), SMC03 (c), and SMC04 (d) sequences at pH **7.0** and **5.0**. The experimental conditions were 0.3 mM DNA, 100 mM KCl, 20 mM phosphate or acetate buffer, 5 °C.

- Despite the apparent complex sequence, the results showed that the wild sequence may form a relatively stable and homogeneous unimolecular i-motif structure, both in terms of pH or temperature.
- The model ligand TMPyP4 destabilizes the structure, whereas the presence of 20% (w/v) PEG200 stabilized it slightly.









mfold prediction for SMC01 sequence

- NMR signals around 15.5 ppm indicate the existence of C·C⁺ base pairs **at pH 5.0**.
- At pH 7.0, signals between 12 and 14 ppm indicate Watson-Crick base pairs, in agreement with mfold [4] predictions.

4. Thermal stability



(a) Fraction of folded DNA
calculated from the absorbance
trace at 295 nm. (b) Plot of
determined T_m values *vs.* pH.

The experimental conditions were 2 mM DNA, 150 mM KCl, 20 mM acetate buffer, pH 5.2.

	рН 4.7				pH 5.2			
Sequence	T _m	ΔH^0	ΔS^0	ΔG^{0}_{370C}	T _m	ΔH^0	ΔS^0	ΔG^{0}_{370C}
	(°C)	(kcal·mol⁻¹)	(cal·K ⁻¹ ·mol ⁻¹)	(kcal·mol⁻¹)	(°C)	(kcal·mol⁻¹)	(cal∙K⁻¹ · mol⁻¹)	(kcal∙mol⁻¹)
SMC01	45	-40.5	-127.1	-1.0	38	-39.0	-124.9	-0.3
SMC02	33 / 44	n.d.	n.d.	n.d.	27 / 42	n.d.	n.d.	n.d.
SMC03	41	-27.9	-88.9	-0.3	36	-30.7	-99.5	0.1
SMC04	49	-26.1	-80.7	-1.0	36	-32.6	-105.7	0.1

3. Molecularity



PAGE of SMC01 and variants in native conditions at pH 8.3 (A) and 5.0 (B). The oligonucleotides sequences are reported in next Table. The average migration of each oligonucleotide *versus*. nucleotide size is reported on the right from two independent experiments including Poly-d(T)45, 30, 24 and 21 markers. The line between points was fitted using a linear regression model.



- PAGE and SEC experiments show that all sequences, apart from SMC02, form rather homogeneous i-motif structures at pH 5.
- SMC02 seems to form both, monomeric and dimeric i-motif structures



- Analysis of thermal melting curves confirmed the dimeric structure of SMC02 at higher concentrations.
- In general, thermal stability of these i-motif structure is low

5. Acid-base properties



Using appropriate multivariate methods, it is possible [5]:

- To determine the number of species of conformations present throughout the experiment,
- 2. To quantify their relative concentration (**a**, **c**, **e**, **g**)
- 3. To recover their **pure spectra (b, d, f, h).**

Acid-base titration of SMC01 sequence. (a) Selected experimental CD spectra. (b) Variation with pH of ellipticity values measured at 290nm. The experimental conditions were 2 mM DNA, 150 mM KCl, 25 °C.



6. Interaction with TMPyP4



(a) CD spectra of SMC01 sequence in **absence** and in **presence of TMPyP4**.

(b) CD spectra recorded along the melting of the previous mixture from 15 to 70 °C. Inset shows the fraction of folded DNA in **absence** and in **presence of ligand**.

The experimental conditions were 2 μ M DNA concentration, 4 μ M TMPyP4, 150 mM KCl, 20 mM acetate buffer, pH 5.2, 25 °C.

Determination of the SMC01:TMPyP4 stoichiometry and binding constant. (a) Experimental molecular absorbance data. (b) Calculated distribution diagram. (c) Calculated pure spectra. (d) Experimental (blue circles) and calculated



pH diagrams of species distribution (a, c, e and g) and pure spectra (b, d, f, and h) calculated from the acid-base titrations of the SMC01 (a, b), SMC02 (c, d), SMC03 (e, f) and SMC04 (g, h) sequences after application of multivariate analysis [4]. Blue line: neutral form; green line: i-motif 1; red line: i-motif 2; cyan line:

protonated form.

The **black line** denotes the pH range where i-motif structures predominate.

 Sequences SMC01 and SMC03 produce two different i-motif structures, which differences may rely on different protonation at the loops.

• The i-motifs formed by the SMC01 sequence are stable through a wider pH range.

(green line) absorbance data at 425 nm.

In (b) and (c) free SMC01, free TMPyP4, 1:2 SMC01:TMPyP4 complex, and 1:4 SMC01:TMPyP4 complex. The experiment was carried out at 25 °C, 150 mM KCl, 20 mM acetate buffer, pH 5.2.

Experimental data in (a) were analyzed with a previously described multivariate analysis method that enables the calculation of the binding constants for the proposed model of species, and the corresponding pure spectra [6].

TMPyP4 binds to the SMC01 i-motif structure with two different stoichiometries (1:2 and 1:4). Binding constants are $10^{6.9}$ and $10^{6.1}$ M⁻¹.

References

- [1] Benabou, S.; Aviñó, A.; Eritja, R.; González, C.; Gargallo, R. RSC Advances 2014, 4, 26956-26980
- [2] Zeraati, M.; Langley, D. B.; Schofield, P.; Moye, A.L.; Rouet, R.; Hughes, W. E.; Bryan, T. M.; Dinger, M. E.; Daniel Christ, C. *Nat. Chem.* **2018**, 10, 631
 [3] Kendrick, S.; Akiyama, Y.; Hecht, S. M.; Hurley, L. H. *J. Am. Chem. Soc.* **2009**, 131, 17667–17676

[4] Zuker, M. Nucleic Acids Res. 2003, 31, 3406-3415
[5] Jaumot, J.; Eritja, R.; Tauler, R.; Gargallo, R. Nucleic Acids Res. 2006, 34, 206-216
[6] Benabou, S.; Aviñó, Lyonnais, S.; González, C.; Eritia, R.; de Juan, A.;

[6] Benabou, S.; Aviñó, Lyonnais, S.; González, C.; Eritja, R.; de Juan, A.; Gargallo, R. Biochimie 2017, 140, 20-33

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The poster background shows the CH⁺·C base pair, the building block of the i-motif DNA.